Sampling and processing

Benthic inveretebrates

Benthic invertebrates



- most frequently used biological quality element (BQE) for monitoring and ES assessment purposes
- reflect long-term conditions, overall degradation
- inhabiting the river bed (visible, up to 0,5 mm)



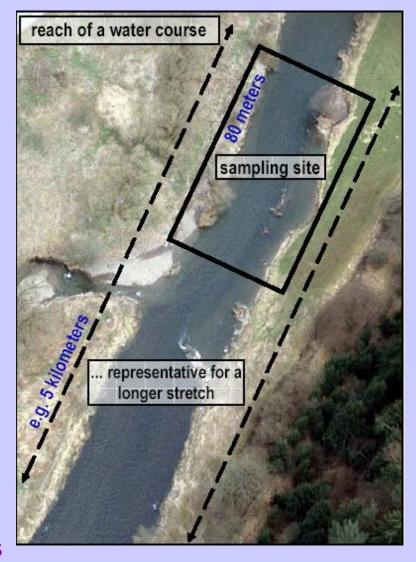
- Assessment of benthic invertebrates Included in STN 757715 Biological analysis of surface water
- quantitative "multihabitat" sampling (AQEM), subsampling, sorting, selection
- quantitative and qualitative assessment
- **Objective:** assessment of water body ES
- standardized sampling method for biomonitoring
- **Result:** list of taxa + density per bed-surface 1,25 m²

Selection of proper sampling site

•representativeness for given reach

assessed stretch:
small streams: 100m
medium and large rivers: 500 m

 sampling – depends on size and depth
 <u>Wadable rivers</u> – pro-rata Multi-Habitat sampling (hand-net)- stony substrate, shallow water (STN EN 16150)
 Selection of sampling methods and devices (STN EN ISO 10870)



Multi-habitat sampling (wadable rivers)

Selection of sampling site

boulder-chute and rocky submerged dyke – not representative

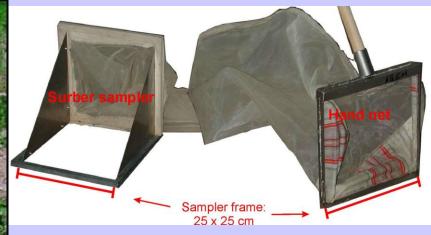


Selection of sampling site



Sampling equipment and devices





 hand-net: 25 x 25 cm, mesh size 500 μm

• GPS, camera, polaroid glasses, preservative



formaldehyd

Multi-habitat estimation

- Substrate assessment
 - ! Correct from stream-bank!
 (stretch of 100m)
- Sampling on substrates with minim. 5% coverage (record in protocol):
- surface area: assessed bed / substrates

Microhabitats recognition !

- avoid high water level, floods, extremely dry period ...
- The best period spring (autumn)

Mineral microhabitats



megalithal (> 40 cm) blocks



macrolithal (20-40 cm) cobbles



mesolithal (6-20 cm) coarse gravel



microlithal (2-6 cm) medium gravel akal (0,2-2 cm) fine gravel

psammal (6 µm-2 mm) sand

Organic microhabitats I.



filamentous algae

submerged macrophytes

emergent macrophytes



living parts of terrestrial plants floating riparian vegetation

fine roots

Organic microhabitats II.



xylal (large wood: trunks, branches)

CPOM - Coarse Partic. Organic Matter FPOM - Fine Partic. Organic Matter



sewage bacteria

debris within the splash zone

Microhabitat estimation

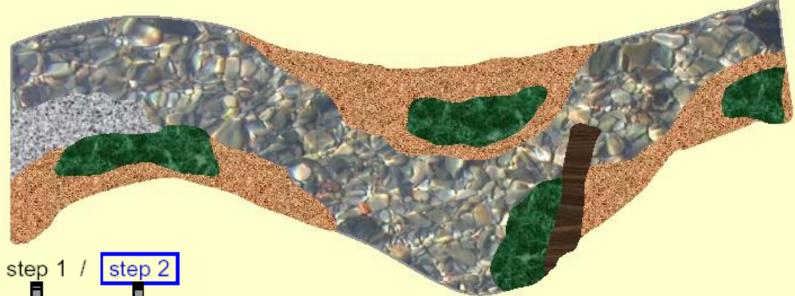


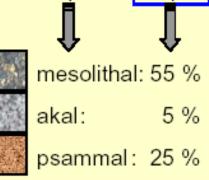


algae

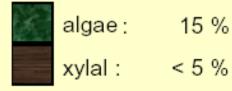
xylal

Microhabitat estimation

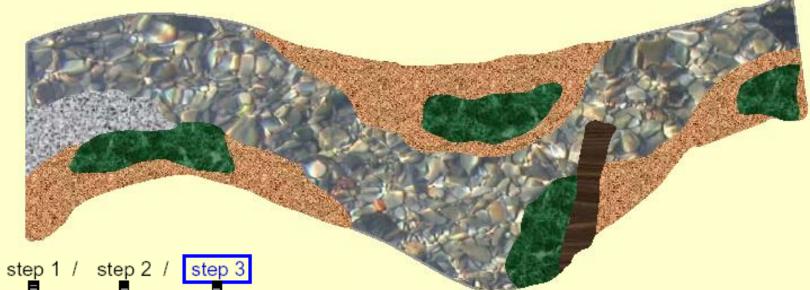


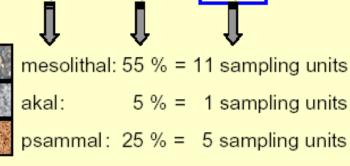


 step: designation of microhabitats
 step: estimation of macrohabitats ratio (< 5 % not considered)



Microhabitat estimation







- algae : 15 % = 3 sampling units
 - < 5 % = <u>no</u> sampling units

- **1.** step: designation of microhabitats
- 2. step: estimation of microhabitats ratio (< 5 % not considered)
- 3. step: setting of sampling units number from considered microhabitats

(20 in total, 1 sampling unit = 5 %)

Microhabitat estimation

- Percentage of estimated habitat rounded to 5%
- Each 5 % = 1 sampling unit

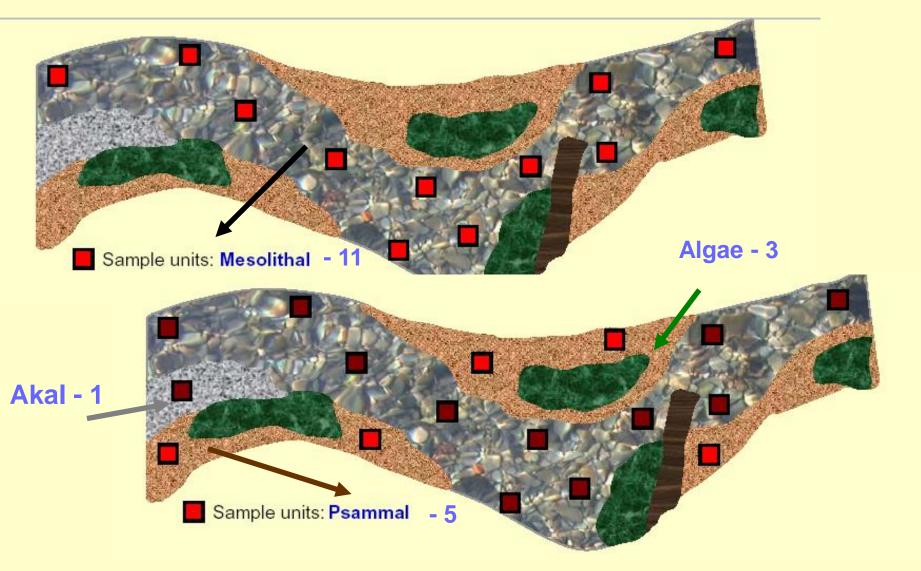
Considering riffles /pools

-

Microhabitats of coverage < 5 % marked by X

	site protoc	ol to define the sa	imple uni	its	ng o	m
	macroinvertebrate multi-habitat-sampling					
	site name	date	investigato	r		
	data in 5% steps, ma	ark the occurrence of rare n	nicrohabitats	with "x"		
MIN	MINERAL SUBSTRATES			no. of sampling units	remat	tks
	megalithal (≻40 cm) upper sides of large cobbles, boulders and blocks, bedrock					
coars	makrolithal (> 20 cm - 40 cm) coarse blocks, head-sized cobbles, with a variable percentage of pebbles, gravel and sand			➡		
	mesolithal (≻6 cm - 20 cm) fist to hand-sized cobbles, with a variable percentage of gravel and sand			11		
coars	m ikrolith al (> 2 cm - 6 cm) coarse aravel (size of a bioson eau to child's fist), with variable percentages of medium to line gravel					
	akal (≻0,2 cm - 2 cm) fine to medium-sized gravel			1		
	psammal / psammopelal (> 6 µm - 2 mm) sand and mud			5		
	argyllal (< 6 μm) sit, loam, clay (inorganic)					
	tech nol (thal 1 (artificial substrates) riprap, stones plastering with major interstices					
	nol thal 2 (artificial substrates) es plastering without interstices, conce	ala				
BIOTIC SUBSTRATES						
alga filam	lgae Iamenious algae, algai tufts		15 %	3		
subr mace	nerged macrophytes ophytes, including moss and Charace	10				
erne e.g.	rgent macrophytes Typha, Carox, Phragmitos					
	g parts of terrestrial plants oots, floating riparian vegetation					
	l (Wood) runks, dead wood, branches, roots		x	0		
CPO depœ	M sits of coarse particulate organic matte	r (e er allen kaves)				
FPO depœ	M sits of fine particulate organic, watter					
sewa sowa (e. g.	age bacteria and stongi and sap ge bacteria and stongi, (Spheerotiks, Beggiatos shilothrir), sludge	robel <i>Leptomitus</i>), sulfur bacteria				
	ic and inorganic matter deposited with motion and changing water levels (e.					
		sum	100%	20		

Multihabitat sampling



Sampling







• kicking = disturbing of substrate

(by foot, hand, brush, screwdriver ...)

- max depth up to 15 cm
- net content empty several times during sampling (to avoid damage of the net)

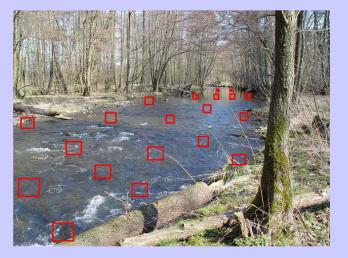
Sampling

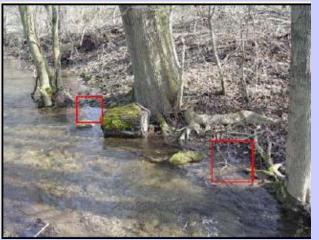
CONSIDER ! Ratio of riffles and pools

By low flow velocity:

• sampling of fine mineral habitats –2-5 cm surface layer directly into the net

•flow artificially generated by hand





Sampling of macrophytes

- bed surface 25 x 25 cm area + corresponding water column
- Organisms are washed down from sampled vegetation (by means of hand-net)



- Mineral substrate from corresponding buttom included as well
- Floating vegetation sampled together with mineral substrate of bottom
- sampling of dominant vegetation



Large rivers (+ deep medium, non - wadable)





- depth and turbidity (estimation ???)
- representative stretch up to 500 m
- -Substrates estimation only in riparian zone / stream-banks
- modified quantitative deep water sampler
 different sampling area
- By homogenous substrate sufficient 10 sampling units

Modified quantitative deep water samplers

Modified sampling method and gear

Standard hand net (Shovel sampler, 25 x 25 cm)

Grab sampler (e. g., Van Veen, 25 x 25 cm, if possible)

(Dredge)

(Core sampler)





sampling area 225 cm² (15cm x 15cm)

Príklady hĺbkových odberových zariadení



Core sampler

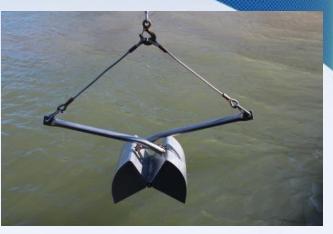


Air-lift sampler



Birge-Ekman grab

Dredge



Van Veen grab



Sample processing

(reduction, selection, conservation, sub-sampling)

Reduction of sampled material



Field selection of organisms

 protected and endangered species, which should be returned into the stream after determination (Astacus astacus, Margaritifera margaritifera, Unio crassus...)

 large and fragile individuals, which should be demaged during the transportation

• rare species of low abundance (*Perla, Perlodes, Ephemera, Anodonta, Unio*)

• max. 20 ind.



Conservation

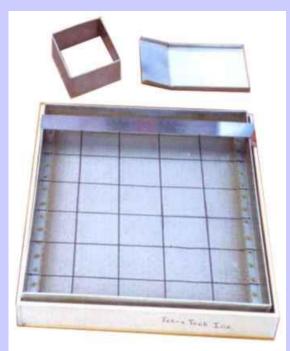
- Formaldehyd (38 %) final concentration 4% (proportion 1:9)
- 95 % ethanol selected ind./species
- In ethanol stored organisms determination features demaged – complicated identification
- Formaldehyd smell removed by washing before further processing (round sieve)

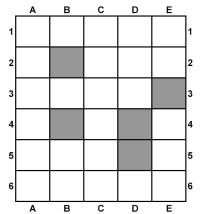


Sub-sampling

SUBSAMPLER

- •30 x 36 cm, rustless metal net (500 µm)
- •split up into 5 x 6 square fields (30 in total)
- •random selection of 5 square fields for sorting







Sub-sampling

sample in subsampler
put in water
homogenized
put out of water
random selection of 5 square fields
(square frame, spoon, scissors, tweezers)



Sub-sampling

•preliminary estimation of individuals number – cca. 500 ± 20%

- <u>if more</u> in 5 square fields another subsampling (sediment again put to subsampler)
- if less continuation (more square fields sorted) untill cca. $500 \pm 20\%$ ind.

PRINCIPLE:

• Min. 1/6 of entire sample (5 square fields

– analyzed ! (record in protocol !)

- Min. 500 organisms! / entire sample
- each square field must be completely collected



Sample analysis and results expression



- Sorting to the systematic groups
- Identification of immediately determinable taxa
- Other and questionable / uncertain taxa
 - conservate by 70% ethanol
- Determination of taxa (permanent slides) + quantitative data

Result: taxa list + density /abundance per 1,25 m²

= Starting point for ES evaluation !!!

Assessment of ES based on benthic invertebrates

Starting point:

• qualitative and quantitative analyses of benthic invertebrates (taxa list + abundance/density)

Calculation of metrics - <u>multimetric index</u> (information on overall ecological status)



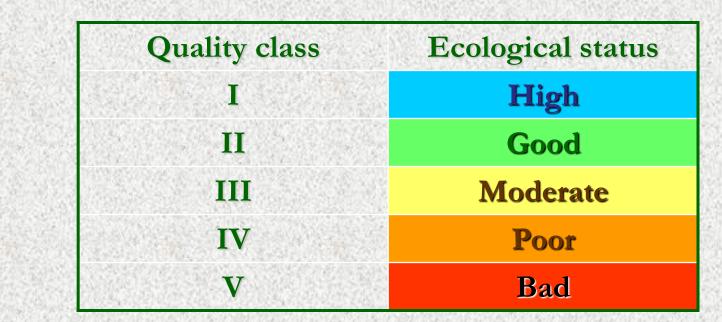
Metrics /indices reflect the autecological characteristics of organisms

> 8 metrics - small streams 11 metrics - medium streams 8 metrics - large rivers (6 - Danube)

Assessment of ES based on benthic invertebrates

Detection of different stressor:

- organic pollution
- degradation of morphology
- general degradation



Thanks for attention

Hydrobiologický kurz, VÚVH, 2007